

Cracow University of Technology  
Faculty of Chemical Engineering and Technology  
Department of Biotechnology and Physical Chemistry



Politechnika Krakowska  
im. Tadeusza Kościuszki

*Microwave-assisted synthesis and  
characterisation of biomaterials based on  
chitosan*

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DR INŻ. MAREK PIĄTKOWSKI

# The Team

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Assistant professor at Cracow University of Technology



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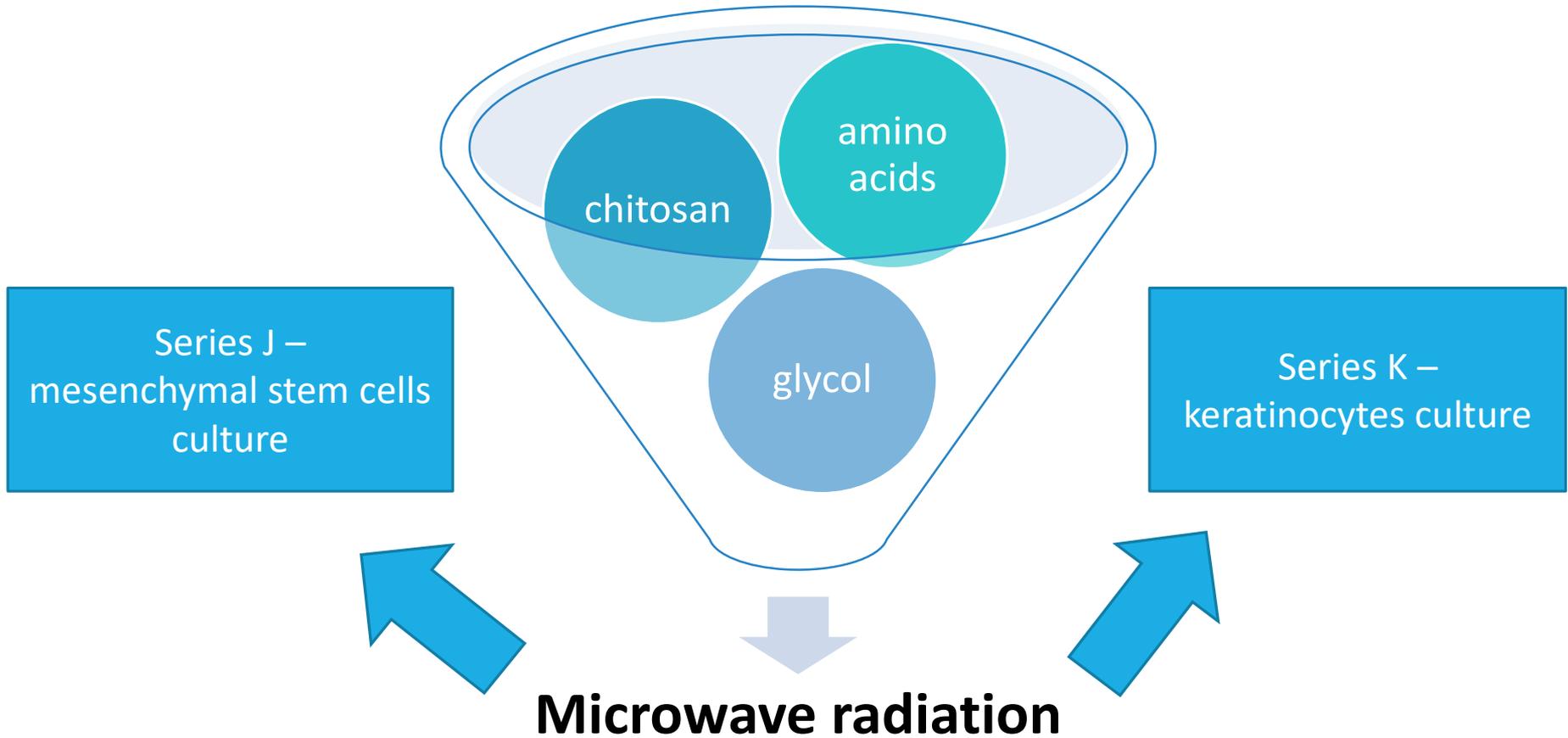
## **Łukasz Janus, MSc Eng.**

Specialist on biotechnology and Green Chemistry  
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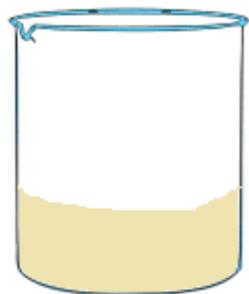




# Synthesis



# Synthesis



Reaction mixture



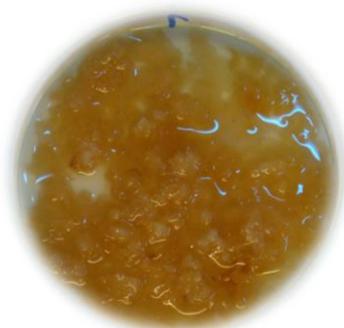
Reaction vessel



Prolabo Synthwave 402  
microwave reactor



**Purification**



Obtained hydrogel

# Synthesis parameters



Tab. 1. Synthesis parameters for Series J

Sample	Asp : Glu : chitosan [g]	Glycol [ml]	Time [min]	Power [W]	Temp. [°C]
J1	0,1 : 0,1 : 1,0	6	30	300	150
J2	0,1 : 0,1 : 1,0	6	30	500	180
J3	0,2 : 0,1 : 1,0	6	30	300	150
J4	0,2 : 0,1 : 1,0	6	30	500	180
J5	0,1 : 0,2 : 1,0	6	30	300	150
J6	0,1 : 0,2 : 1,0	6	30	500	180
J7	0,2 : 0,2 : 1,0	6	30	300	150
J8	0,2 : 0,2 : 1,0	6	30	500	180
J9	0,3 : 0,3 : 1,0	6	30	300	150
J10	0,3 : 0,3 : 1,0	6	30	500	180



Steam sterilisation



EO sterilisation

# Synthesis parameters



Tab. 2. Synthesis parameters for Series K

Sample	Asp : Glu : chitosan [g]	Glycol [ml]	Time [min]	Power [W]	Temp. [°C]
K58	0,10 : 0,20 : 1,00	10	25	300	150
K59	0,10 : 0,20 : 1,00	10	25	500	180
K60	0,20 : 0,10 : 1,00	10	25	300	150
K61	0,20 : 0,10 : 1,00	10	25	500	180
K62	0,20 : 0,20 : 1,00	10	25	300	150
K63	0,20 : 0,20 : 1,00	10	25	500	180
K64	0,25 : 0,20 : 1,00	10	25	300	150
K65	0,25 : 0,20 : 1,00	10	25	500	180
K66	0,25 : 0,25 : 1,00	10	25	300	150
K67	0,25 : 0,25 : 1,00	10	25	500	180

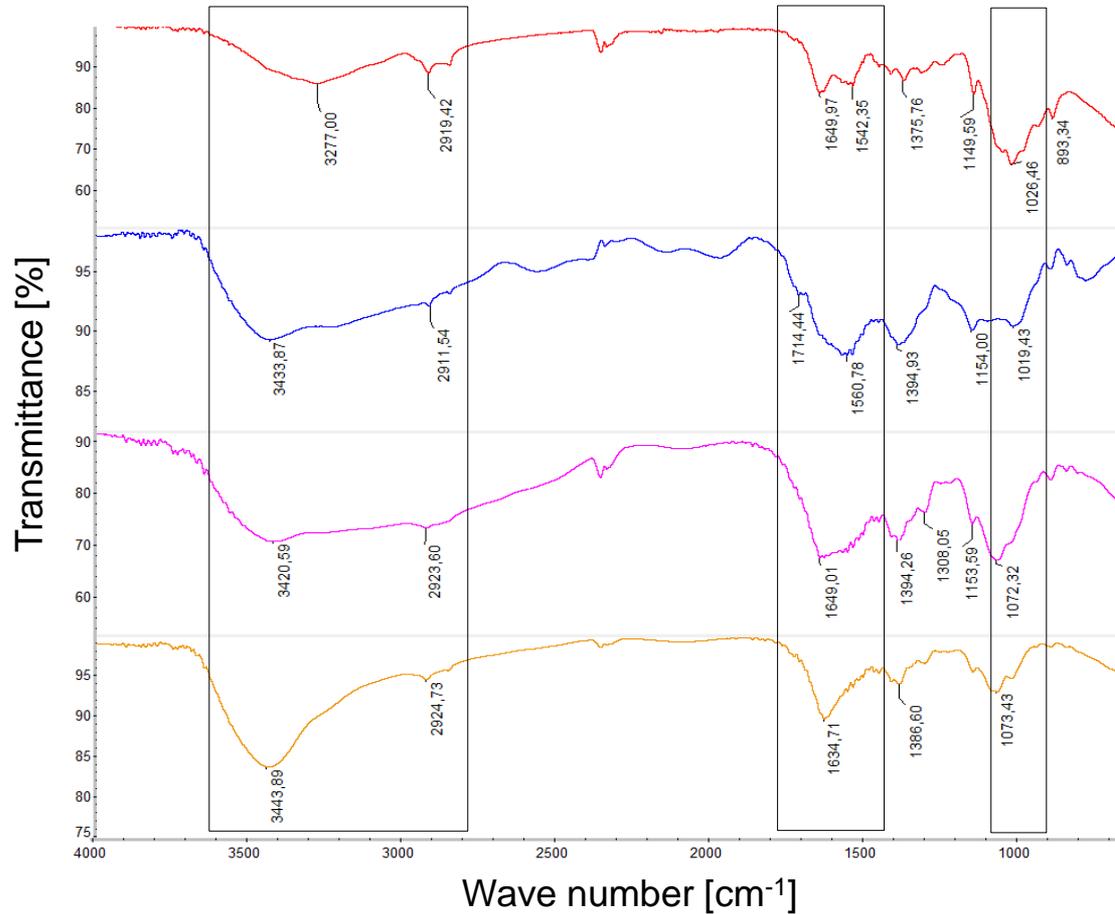


Gamma radiation  
sterilisation (25 kGy)



Gamma radiation  
sterilisation (35 kGy)

# FTIR analysis - Series J



Chitosan



Scaffold



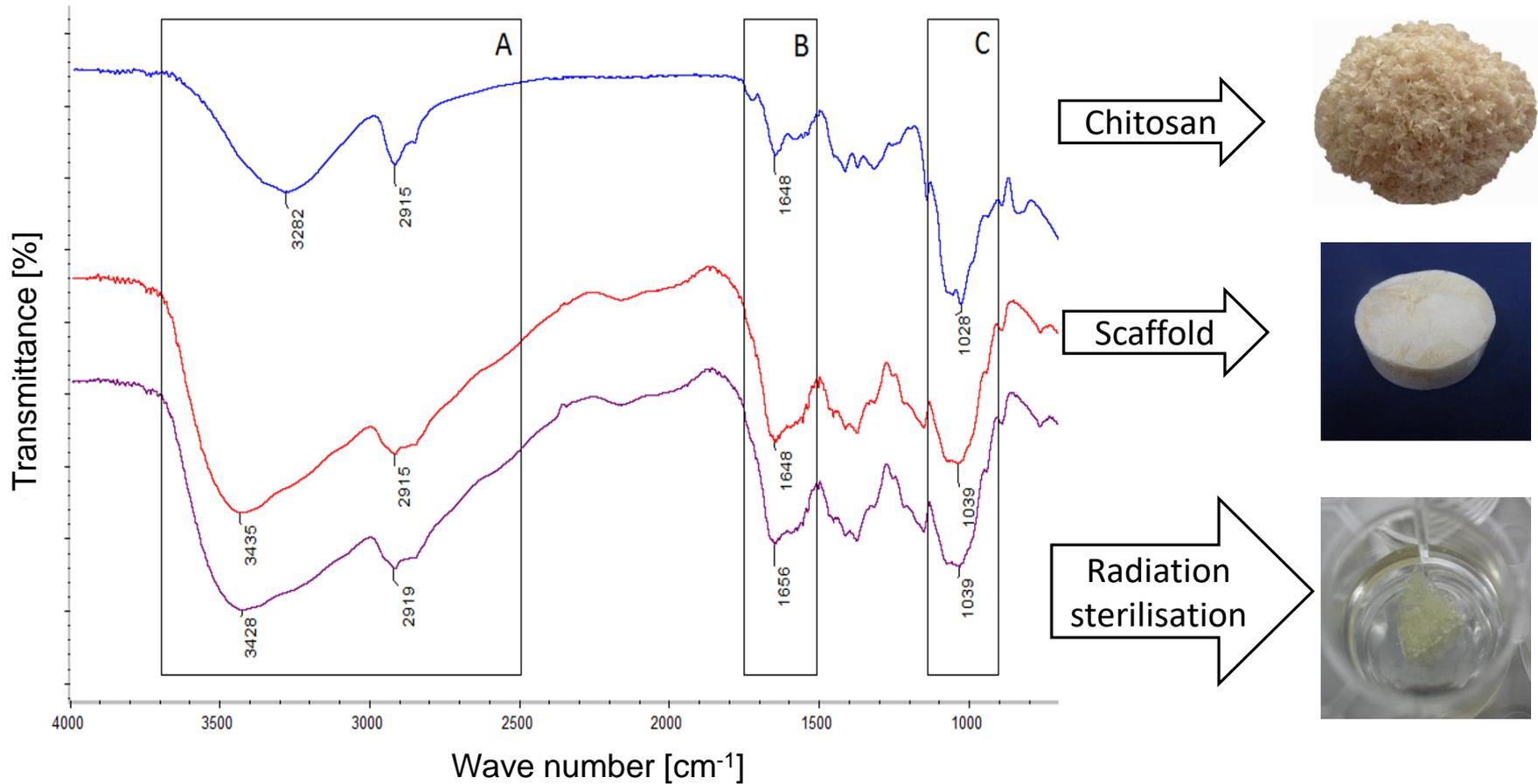
Thermal sterilisation



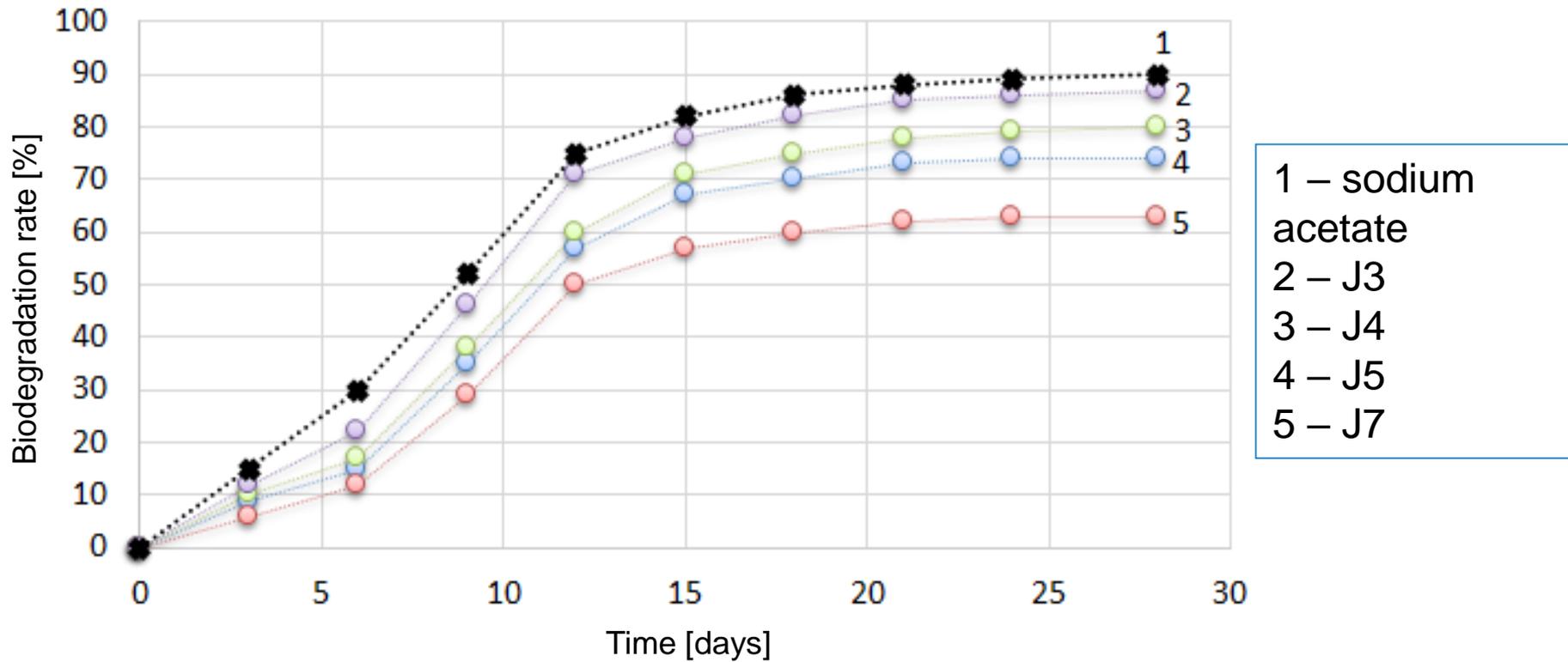
Chemical sterilisation



# FTIR analysis - Series K



# Biodegradation – Series J



# SEM analysis – Series J

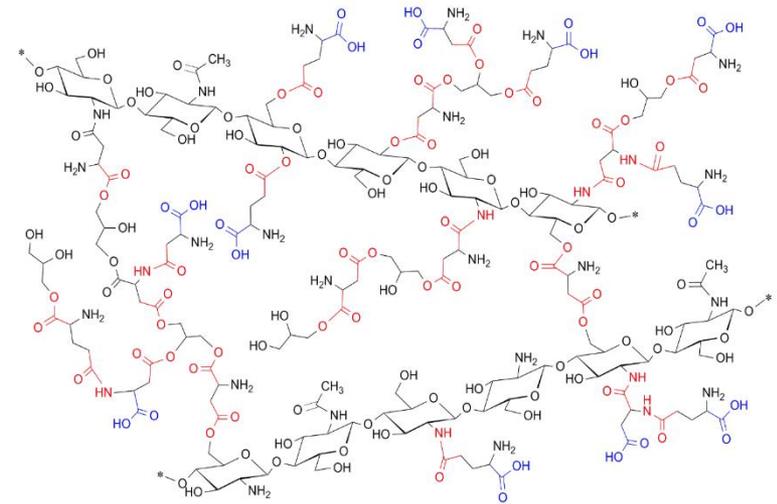
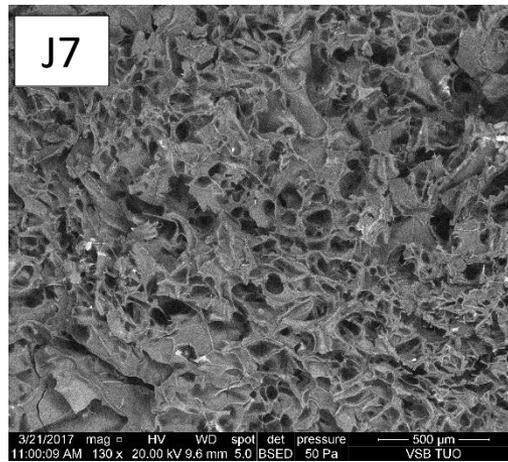
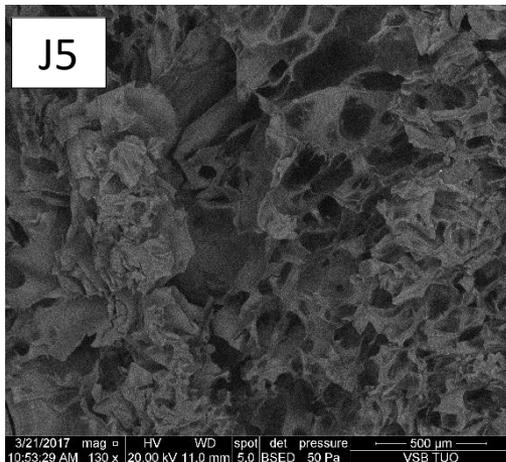
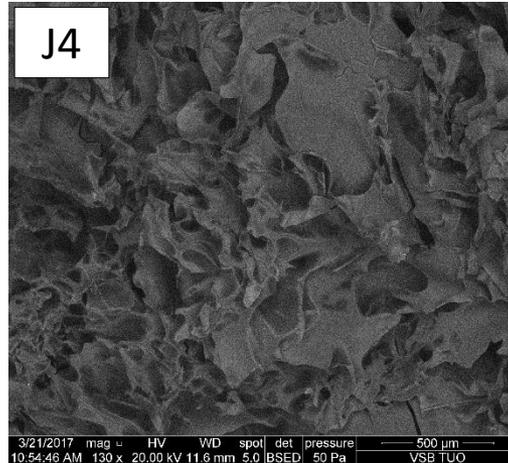
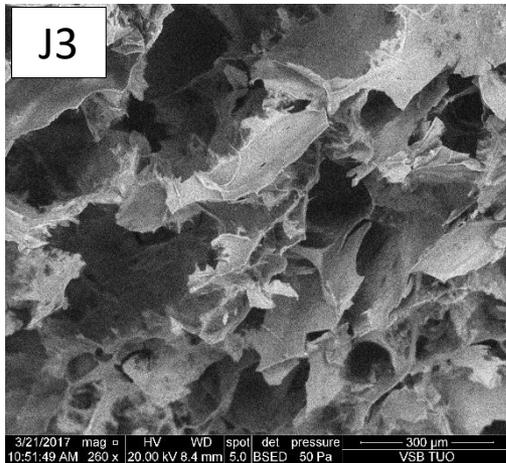


Fig. 2. Proposed chemical structure of crosslinked chitosan

# Antibacterial properties - Series J



Tab. 3. Growth of bacteria on prepared scaffolds

Sample	AmnioGrow	DMEM	KSFM
J3	-	-	-
J4	-	-	-
J5	-	-	-
J7	-	-	-

Both scaffolds before and after sterilization in all tested media were not colonized by bacteria and fungi.

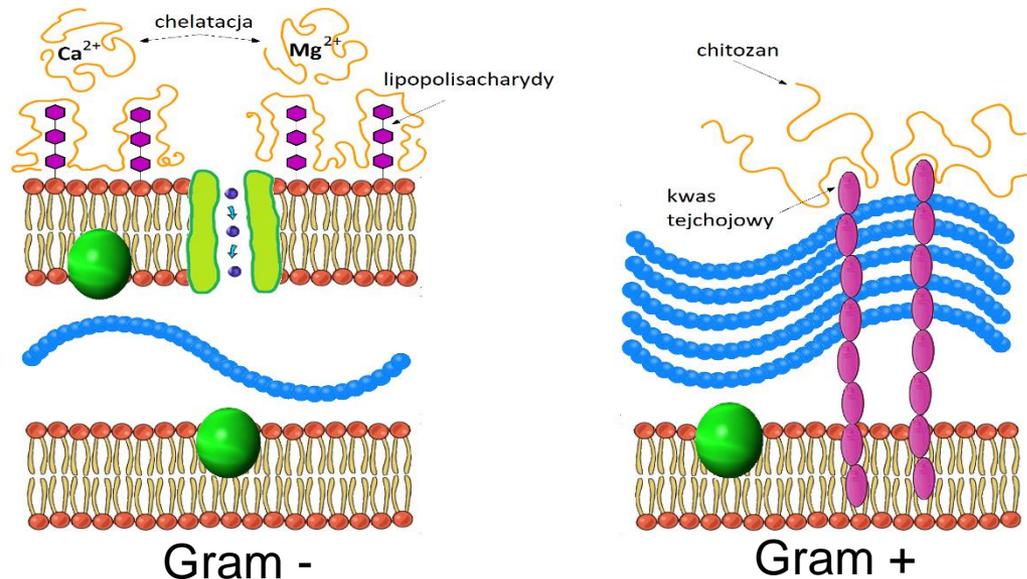
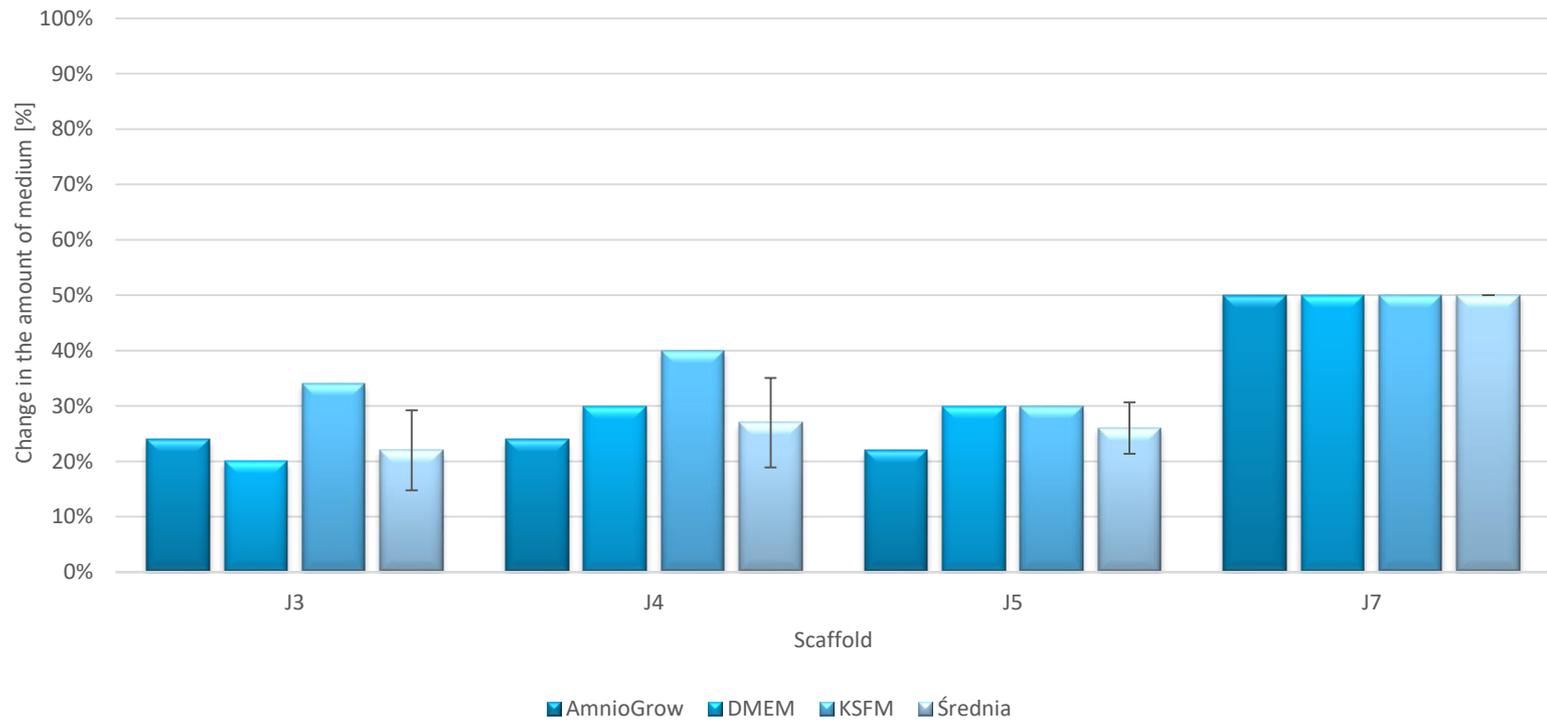


Fig. 3. Mechanism of antibacterial activity of chitosan

# Swelling - Series J



Absorption of culture medium depending on scaffold and type of medium



# Stability in culture media - Series J

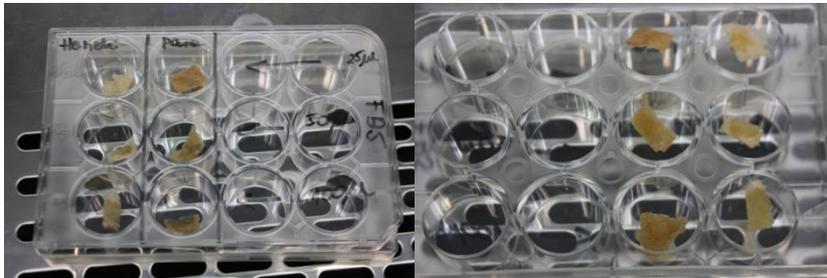


Fig. 4. Attachment to the bottom of the plate (J7)

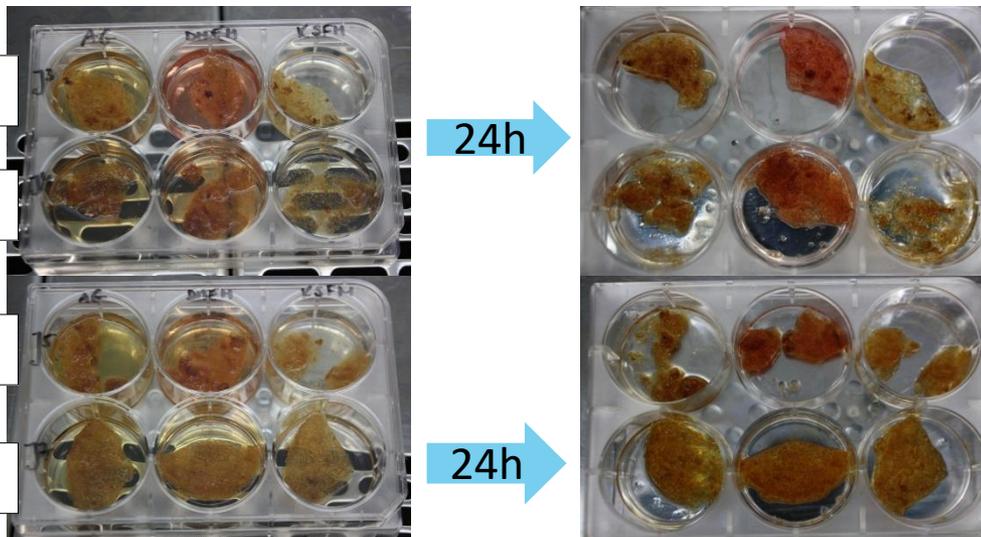
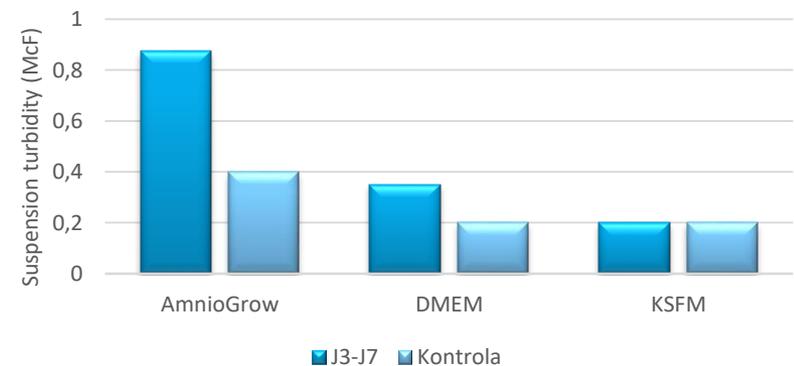


Fig. 5. Swelling in culture media

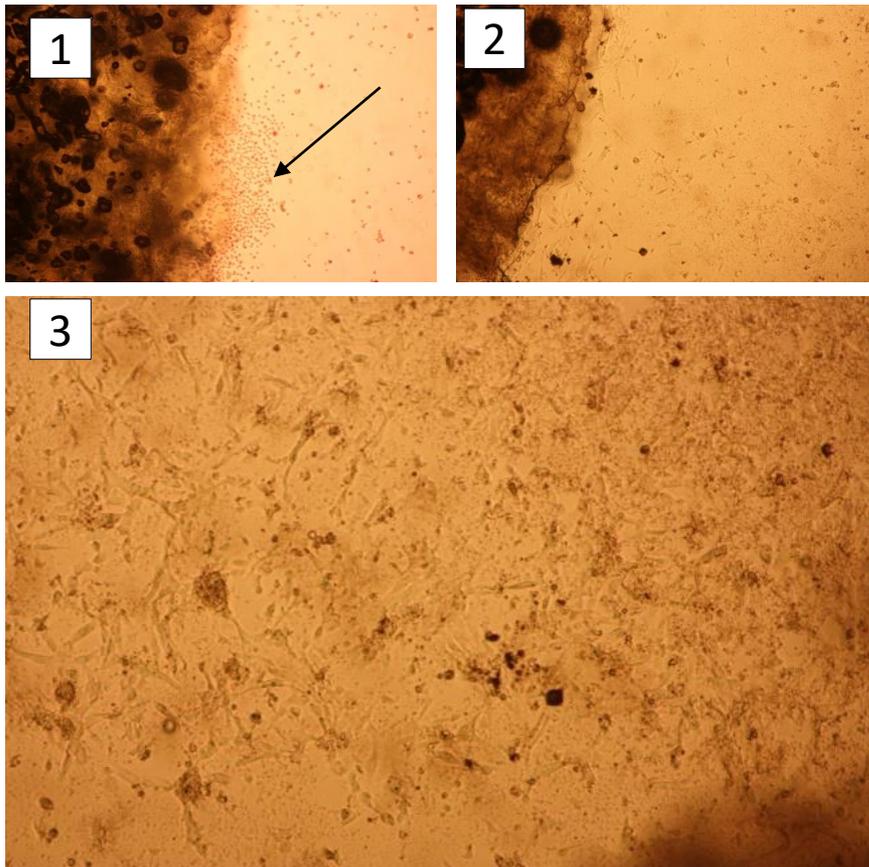
The average suspension turbidity depending on the scaffold



The average suspension turbidity depending on the culture media



# Population doubling - Scaffold J7



Population doubling depending on sterilisation method

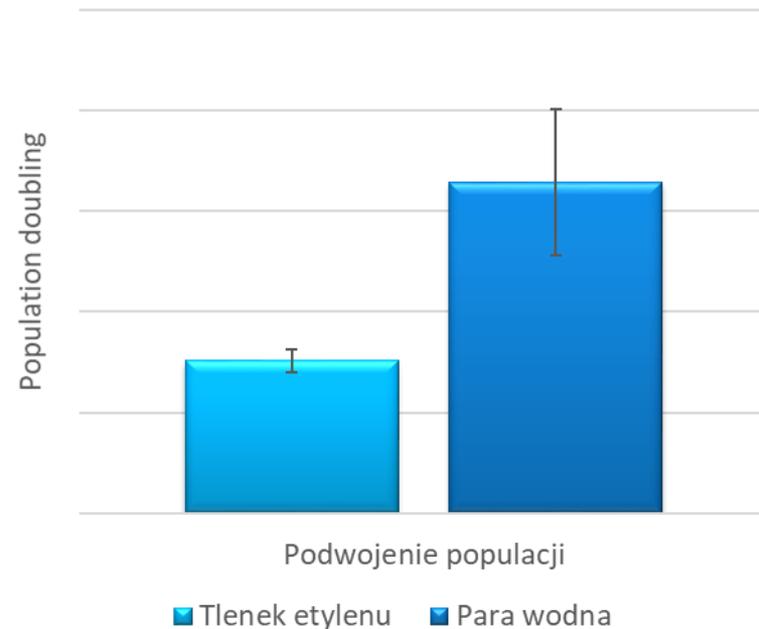
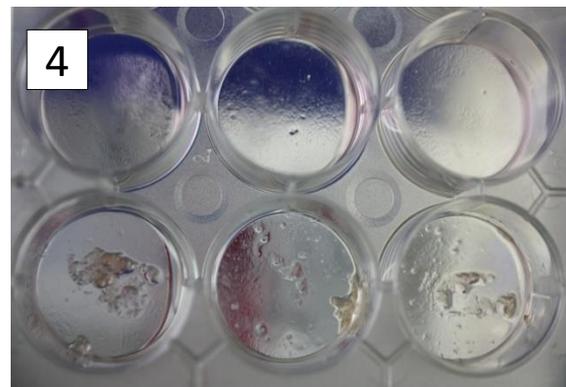
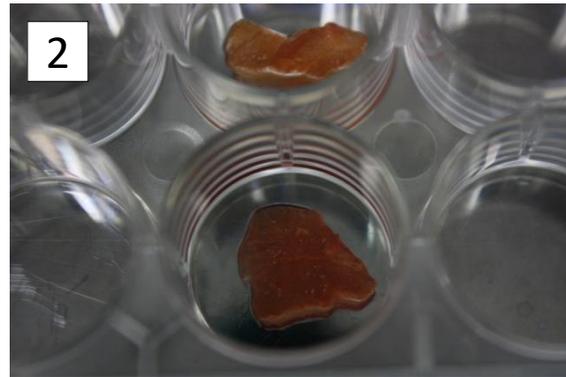
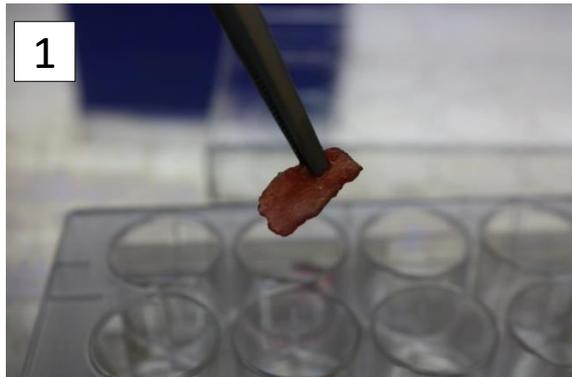


Fig. 6. 1 – Sown cells, 2 – No contact inhibition, cell coating, 3 - Proliferation

# Scaffold stability - J7



1. Manipulation with tweezers doesn't affect the integrity
2. Scaffold have retained its form after the transfer
3. Disintegration of scaffold sterilised with ethylene oxide
4. Fragments of scaffold sterilised with EO permanently attached to the plate

Fig. 7. Stability of the scaffold after cell culture

# Cells settlement

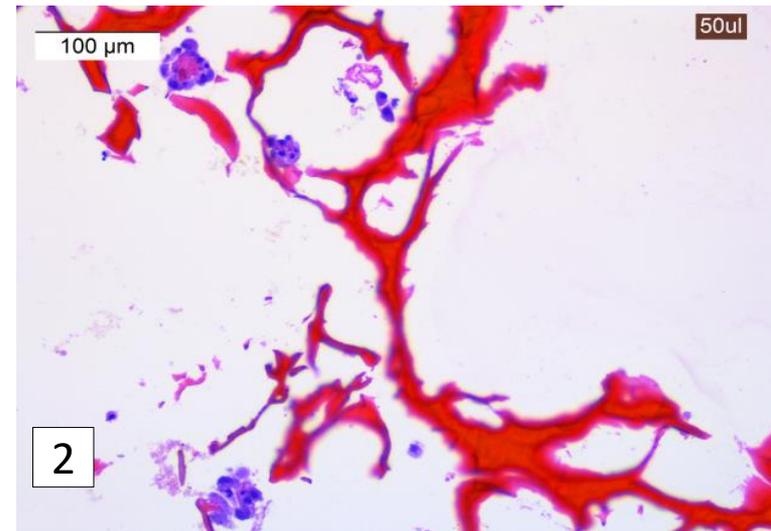
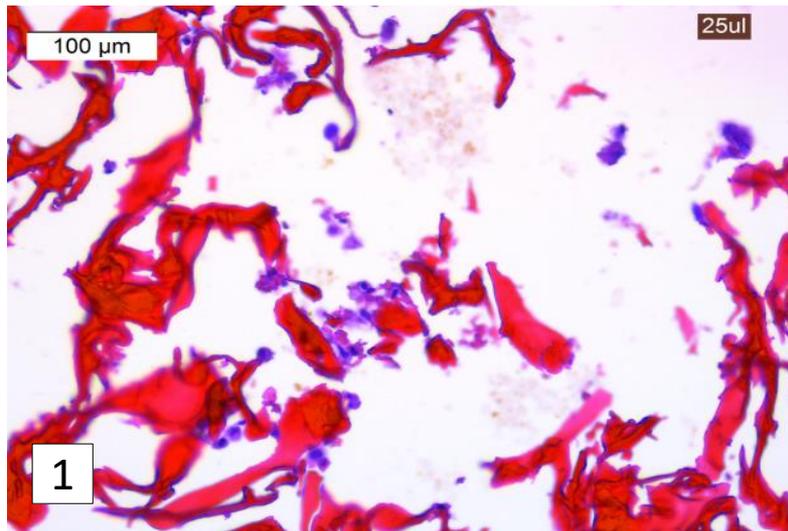
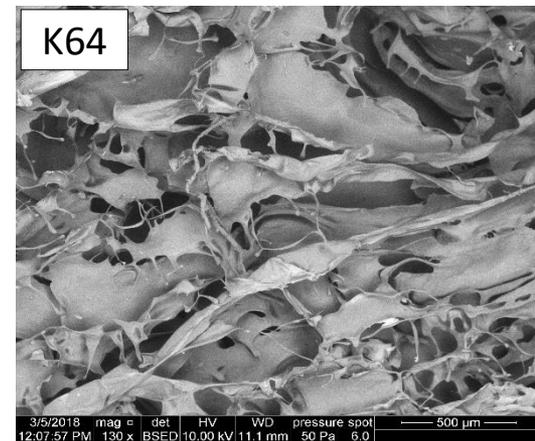
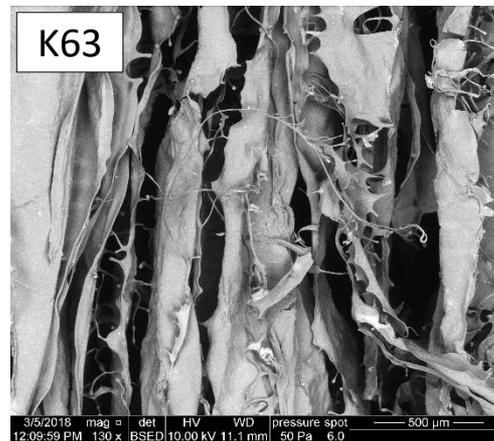
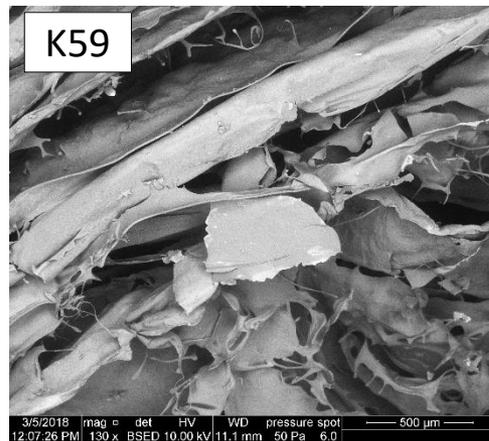
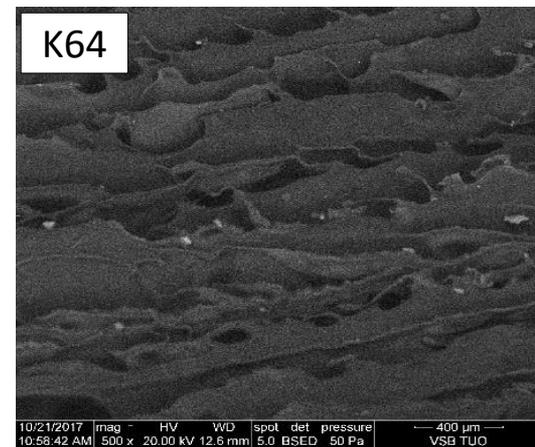
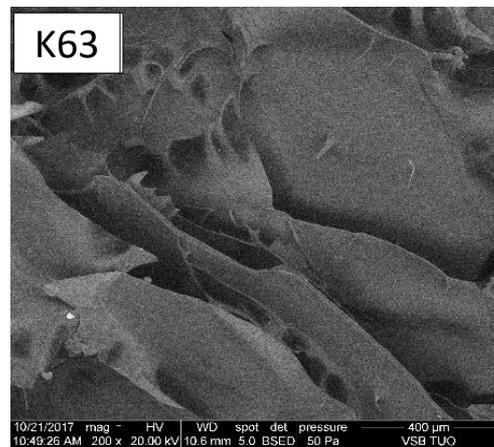
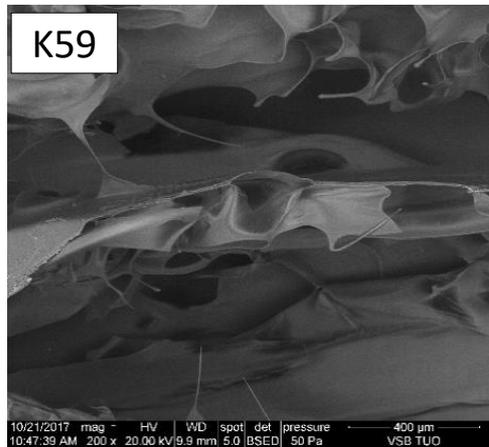


Fig. 8. Histopathological analysis showing the presence of cells in chitosan scaffold, sterilized both with steam (1) and with ethylene oxide (2)

Visible blue-violet stained cells and matrix fibres stained in red

# SEM analysis – Series K



# Scaffold sterility - Series K



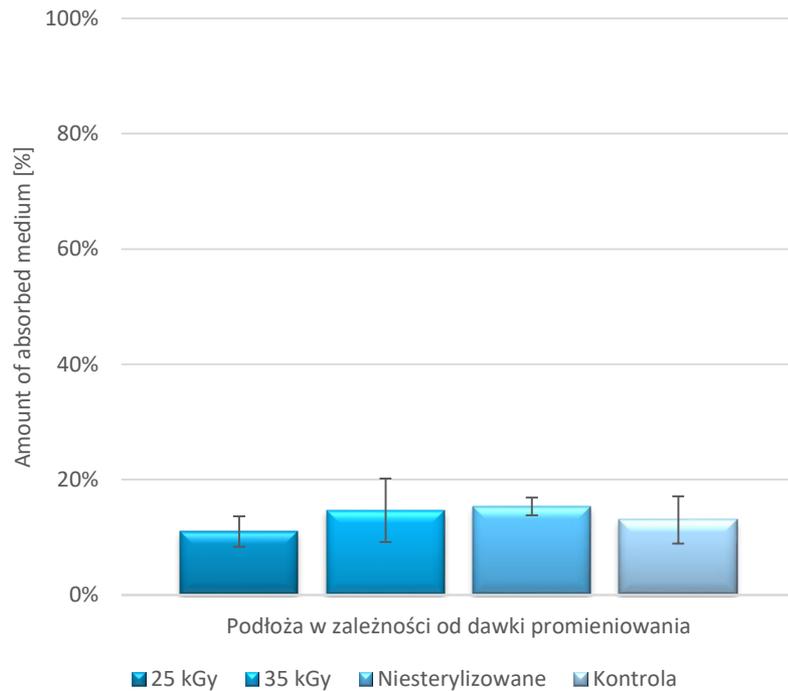
Tab. 4. Growth of bacteria on prepared scaffolds

	K63/ 95°C	K64/ 95°C	K59/ 120°C
Before sterilisation	S. epidermidis MSSE	-	S. epidermidis MRSE
Sterility in culture with antibiotics before radiation	-	-	-
Sterility after irradiation (25 kGy)	-	-	-
Sterility after irradiation (35 kGy)	-	-	-
Sterility in culture without antibiotics after irradiation (25 kGy)	-	-	-
Sterility in culture without antibiotics after irradiation (35 kGy)	-	-	-

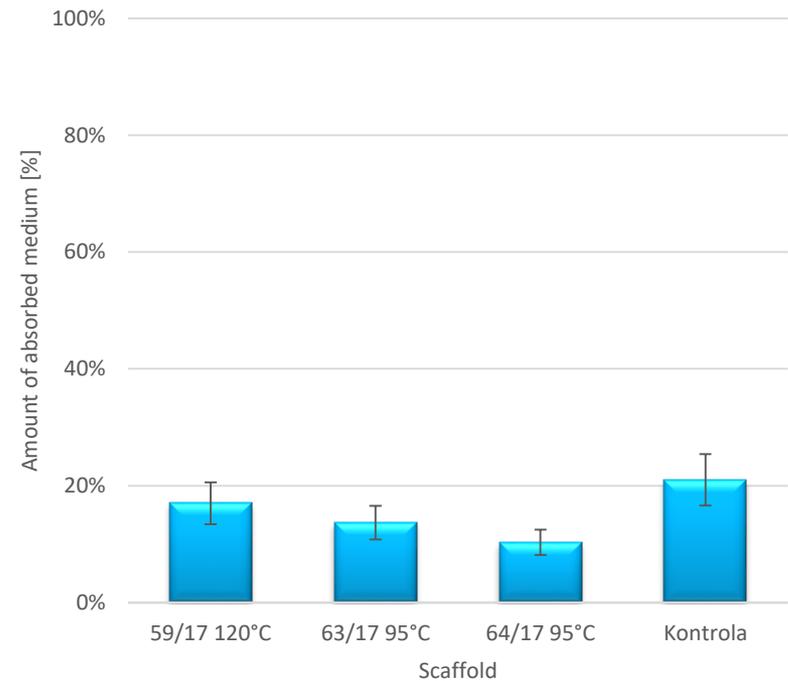
# Swelling - Series K



Amount of absorbed medium depending on sterilisation method



Amount of absorbed medium depending on scaffold



# Cells adherence - Series K



Tab. 5. Percentage expressed number of cells that did not adhere to the bottom of the culture vessel depending on the dose of sterilization radiation and the type of medium

	K59/ 120°C	K63/ 95°C	K64/ 95°C	Control
Unsterile	3,7%	2,9%	5,7%	6,5 %
Sterilised with 25kGy	2,6%	13,6%	6,5%	
Sterilised with 35kGy	14,3%	5,5%	4,1%	
Average	6,8%	7,3%	5,4%	

# Cells adherence - Series K



Cells adherence;  
sterilisation with  
25 kGy after  
96h

Cells adherence;  
sterilisation with  
35 kGy after  
96h

Degree of  
enzymatic  
detachment

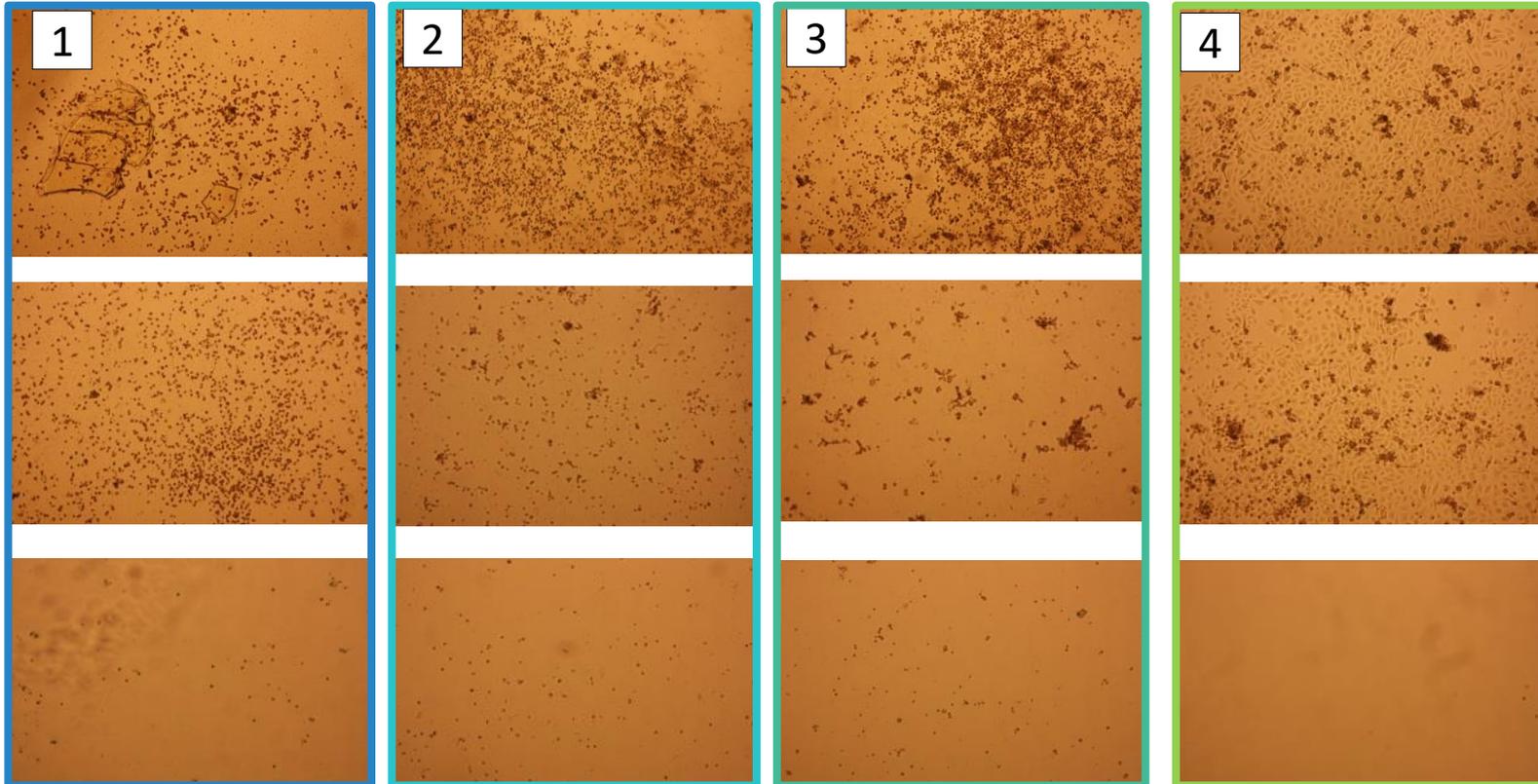
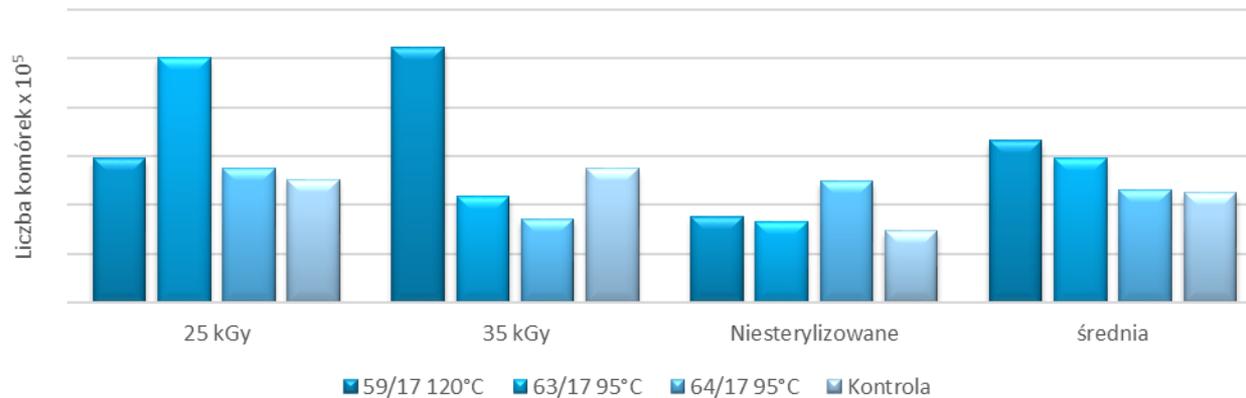


Fig. 9. 1 - Sample K59, 2 - Sample K63 , 3 - Sample K64, 4 - Control

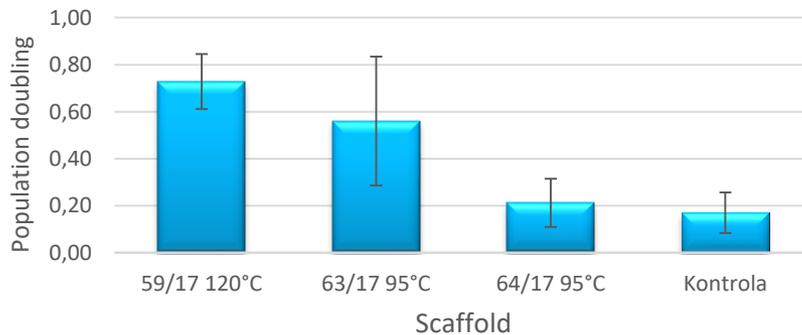
# Population doubling – Series K



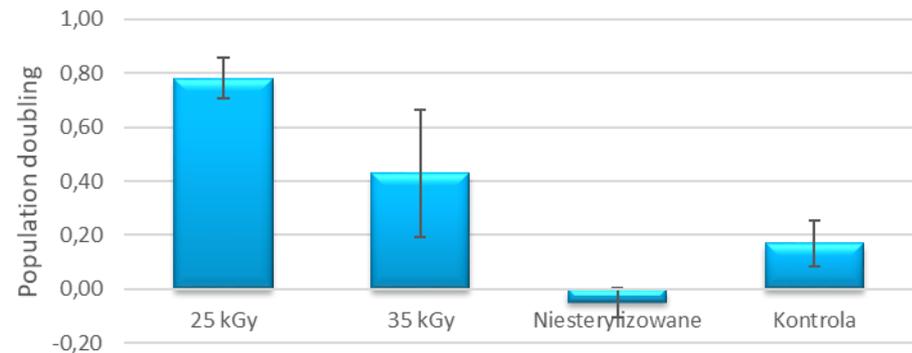
Number of cells after culture depending on scaffold and sterilisation dose



Population doubling depending on scaffold



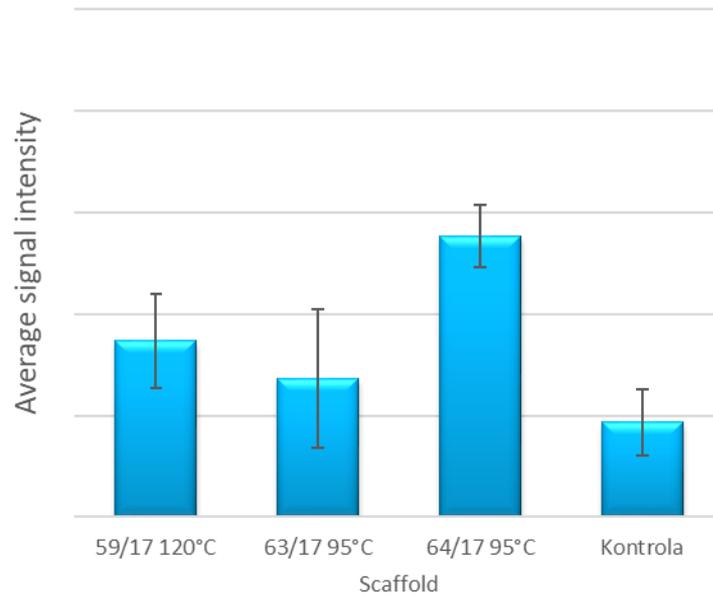
Population doubling depending on sterilisation dose



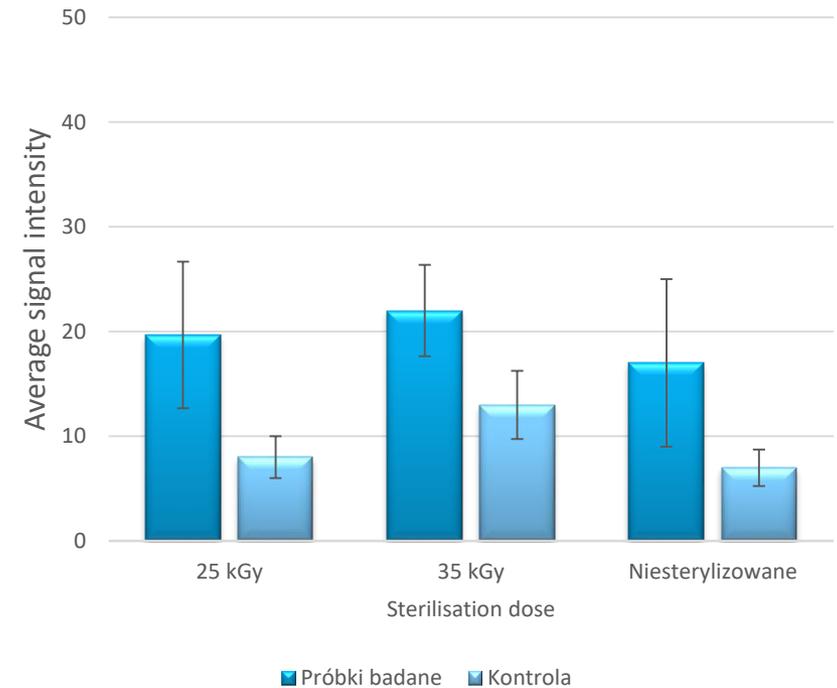
# Reactive Oxygen Species - Series K



Amount of reactive oxygen species (ROS) depending on scaffold



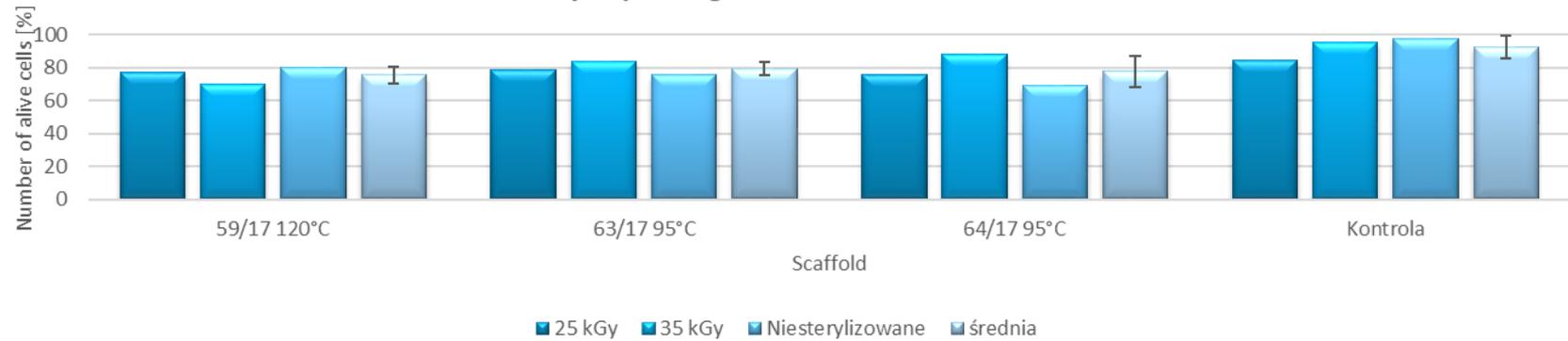
Amount of reactive oxygen species (ROS) depending on sterilisation dose



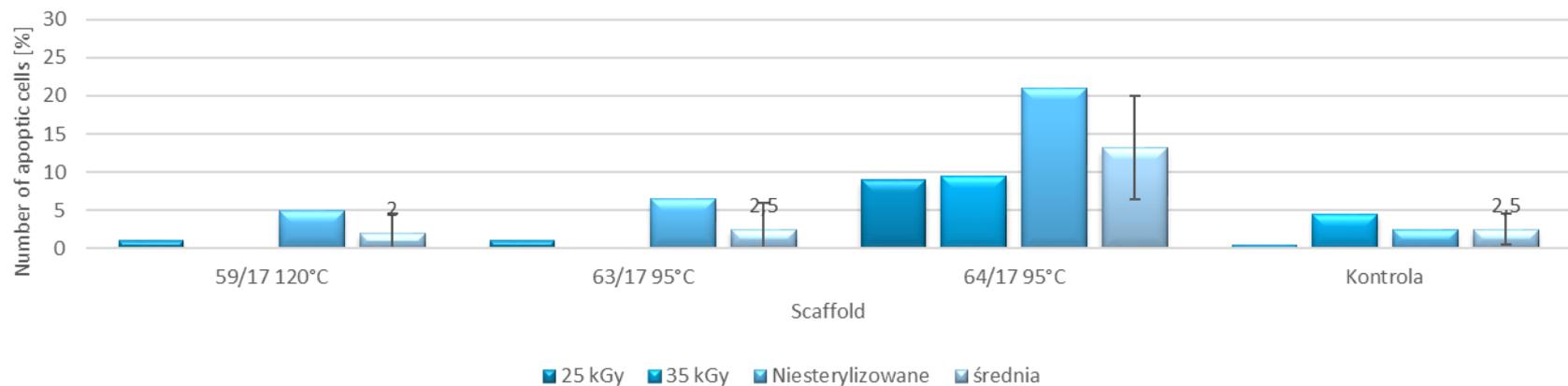
# Cells viability - Series K



Cells viability depending on scaffold and sterilisation dose



Number of cells in apoptic phase depending on scaffold and sterilisation dose





Thank You for Your  
Attention